

**Review****Mutated Genes in Pancreatic Cancer**

Song-bing He, De-chun Li\*

*Department of General Surgery, the First Affiliated Hospital, Soochow University, Suzhou 215006, China***CLC number: R735.9 Document code: A Article ID: 1000-9604(2010)02-0093-06****DOI: 10.1007/s11670-010-0093-9**

© Chinese Anti-Cancer Association and Springer-Verlag Berlin Heidelberg 2010

**ABSTRACT**

Pancreatic cancer continues to be a deadly malignancy with still high mortality and poor survival. Little progress has been made on the treatment of advanced pancreatic cancer despite the significant advances in understanding, diagnosis, and access to conventional and novel treatments. Molecular pathology of the lesion is the key of our understanding of the mechanisms underlying the development of this cancer and will probably help us in earlier diagnosis and better therapeutic results. New treatment strategies and a more careful evaluation of innovative therapies are clearly needed for this disease. In view of many findings pancreatic cancer should be regarded as a systemic disease, and over the last several years, investigators have gained a better understanding of the molecular biology and events that lead to the development of malignancies. We here review the novel developments in the systemic exploring for commonly mutated genes in pancreatic cancer.

**Key words: Pancreatic cancer; Epithelial growth factor; Matrix metalloproteinases; Oncogenes****INTRODUCTION**

Pancreatic cancer (PC) is one of the most lethal cancers worldwide, as indicated by its mortality incidence ratio of 98%<sup>[1]</sup>. The prognosis for pancreatic cancer is extremely poor, and the high mortality can be attributed to the ambiguous symptoms, a distinct lack of early diagnostic methods and a lack of effective therapy<sup>[2]</sup>. Surgery remains the only curative treatment option for this disease but 80% of pancreatic cancer patients have unresectable or metastatic disease<sup>[3,4]</sup>. Moreover, adjuvant chemotherapy or radiotherapy was assessed in several trials in order to improve patients' prognosis<sup>[5,6]</sup>. Thus, most patients receive palliative treatment with the aim of an improved quality of life.

In view of many findings pancreatic cancer should be regarded as a systemic disease, and over the past few decades, knowledge of the genetic defects

involved in tumor formation and growth has increased rapidly. There is an imperative need of analyzing and understanding the primitive lesions that lead to invasive pancreatic cancer. Identification of cancer escape mechanisms and critical cell survival pathways has launched the development of novel antitumor agents. This review will focus on the recent developments in the systemic exploring for commonly mutated genes in pancreatic cancer.

**K-ras**

K-ras activation is known to contribute to cancer cell survival in a number of different tumor types and experimental systems. In mouse models of cancer, somatic activation of oncogenic K-ras is necessary for the early onset of tumors, and its continuous production is necessary for the maintenance of tumor viability<sup>[7]</sup>. Molecular genetic analysis has identified that activating K-ras mutations, as the earliest and most common genetic mutations in pancreas cell transformation and tumor progression, are found not only in 70% to 95% of pancreatic carcinoma tissues but also in pancreatic juice, fine-needle aspirations of

**Received** 2009-12-08; **Accepted** 2010-02-23

This work was supported by the Scientific Research Foundation of Ministry of Public Health of China (No. WKJ20042011).

\*Corresponding author.

E-mail: hesongbing2@yahoo.com.cn

the pancreas, duodenal fluid, and blood and stool of pancreatic cancer patients<sup>[8,9]</sup>. Recent study demonstrated that the introduction of a promoterless full-length K-ras cDNA could efficiently suppress endogenous K-ras gene expression in human pancreatic cancer cells. This inhibition was achieved when the adequate levels of the responsible effectors were reached. Knockdown of K-ras expression in pancreatic cancer cells resulted in slower cell proliferation and lower tumorigenicity in mice<sup>[10]</sup>. Moreover, K-ras mutations have been reported as a negative prognostic factor after surgery and adjuvant chemoradiation in pancreatic cancer<sup>[11,12]</sup>.

### p53

In human cancers, the most frequent mutant gene is p53, which has a central position in cell cycle regulation through its role in inactivating a variety of genes and interrupting cell proliferation at G1/S checkpoint. p53 mutations cause loss of the above functions and are central in carcinogenesis leading to uncontrolled cell growth and proliferation, increased cellular survival and chromosomal instability. It has been reported that p53 mutations are found in up to 76% in pancreatic cancer<sup>[13,14]</sup>. Usually, mutations of p53 are not specific but rather sporadic and are resulting in the production of a mutant p53 protein which lives longer than the wild type(normal) p53 protein. Furthermore, p53 mutations may lead to resistance to chemotherapy treatments due to impaired p53-induced apoptosis<sup>[15]</sup>. There is emerging evidence that the oncogenic potential of human and/or murine double minute-2 protein(hdm2) stems not only from its ability to counteract tumor suppressor p53 but also from its less understood p53-independent functions. Sui X et al concluded from their results that hdm2 is expressed in pancreatic cancer cells as a result of activated Ras signaling, and that it regulates cellular proliferation and the expression of three novel target genes by p53-independent mechanisms<sup>[16]</sup>. However, whether these mutations have also a prognostic significance remains unclear. Few researchers have suggested a shorter overall survival in pancreatic cancer patients carrying p53 mutation compared to p53(-) patients<sup>[13,17]</sup>.

### p16

Mutation and loss of p16 activity results in absence of the inhibitory effects at the level of cyclin D-CDK4 interaction, thereby promoting cell cycle progression. Genetic alterations of this locus through gene mutation, deletion, or promoter hypermethylation are found in 80% to 95% of sporadic pancreatic

cancers<sup>[18]</sup>. Genetic analyses have shown that p16 alterations are very common in pancreatic adenocarcinomas but these alterations are not necessarily seen in cultured cell lines. Moreover, loss of p16 expression could be correlated with less differentiated tumors, shorter overall survival, and the presence of metastatic disease<sup>[19,20]</sup>. The polymorphic genotypes of the p16 gene were associated with significantly shorter time to tumor progression and poorer response to therapy in pancreatic cancer<sup>[21]</sup>. Functional inactivation of the tumour suppressor p16 marks a key event in pancreatic carcinogenesis. Conversely, reconstitution of the biological action of p16 provides an attractive therapeutic approach. Schulz P et al addressed the consequences of p16 re-expression in MiaPaca-2 pancreatic cancer cells in vivo in the orthotopic tissue context. They found p16 capable of reducing primary tumor growth. In addition, p16 restitution resulted in a marked reduction of tumor angiogenesis, largely accounted for by a p16-dependent inhibition of lymphangiogenesis<sup>[21]</sup>.

### DPC4/SMAD4

Another tumor suppressor gene which is involved in pancreatic cancer pathogenesis is the DPC4 gene also known as SMAD4. In normal cells the product of this gene plays a role in TGF- $\beta$  mediated signal transduction, gene transcription and growth arrest<sup>[22]</sup>. Inactivation of DPC4 facilitates uncontrolled cellular growth and proliferation. It is found that about 50% of pancreatic cancers present mutant genes<sup>[24]</sup>. Loss of the DPC4 expression is a rather late event in the pathogenesis of pancreatic cancer, as this gene was expressed normally in PanIN1 and 2 and only in 30% of cases with PanIN3<sup>[23,24]</sup>. Cao D et al utilized the Gene Logic Inc. BioExpress<sup>TM</sup> platform and Affymetrix U133 GeneChip<sup>®</sup> set to determine the changes in gene expression associated with SMAD4 gene inactivation in a series of well-characterized pancreatic cancer cell lines to define the downstream effects associated with SMAD4 gene inactivation, which offers insight into the role played by SMAD4 in pancreatic cancer, as well as providing potential novel targets for the development of molecular therapeutics in pancreatic cancer<sup>[25]</sup>. Pancreatic cancers are represented by distinct genetic subtypes with significantly different patterns of failure. Determinations of DPC4 status at initial diagnosis may be of value in stratifying patients into treatment regimens related to local control versus systemic therapy<sup>[26]</sup>. Whether the DPC4 status has a prognostic value remains controversial, as in a few studies positive DPC4 status was associated with better outcome and survival post resection<sup>[25,27]</sup>,

but in other studies DPC4 expression was associated with worse outcome after surgery or adjuvant chemotherapy<sup>[23,28]</sup>.

### HER-1/EGFR

Changes in the expression of a range of proteins involved in the control of the cell cycle, proliferation, apoptosis, and invasiveness, such as human epidermal growth factor receptor (HER-1/EGFR) play a critical role in the development of pancreatic cancer<sup>[29]</sup>. In pancreatic cancer, amplification of the EGFR gene has been demonstrated by many researchers but the incidence rate ranges from 16% to 65%<sup>[30]</sup>. The coexpression of HER-1/EGFR has been shown to stimulate tumor cell proliferation, and elevated HER-1/EGFR levels are linked to poor disease outcomes and lower sensitivity to chemotherapy<sup>[31]</sup>. The signal activated by EGFR, penetrating into cell nucleus, accelerates protooncogene phosphorylation, and it is therefore assumed that a serious prognosis of pancreatic cancer is also due to high expression of EGFR and its gene polymorphism<sup>[32]</sup>. Increased EGFR gene copy number and elevated EGF levels are present in a significant proportion of pancreatic cancer patients, and this may reflect increased EGFR pathway dependence with improved sensitivity to EGFR-targeted therapy<sup>[33,34]</sup>. Agents that target the HER-1/EGFR pathway are at the forefront of development, and a significant number of ongoing trials continue to explore the potential of these therapies for this disease<sup>[35]</sup>.

### COX-2

Cyclooxygenase(COX)-2 is inducible by cytokines and intracellular signals produced at sites of inflammation, it can also be induced in various normal tissues by the hormones of ovulation and pregnancy, growth factors, oncogenes and tumor promoters. Experimental data regarding COX-2 biology have also accumulated in pancreatic cancer<sup>[36]</sup>. Preclinical studies testing the COX-2 expression in pancreatic cancer cells in comparison to PanINs, chronic pancreatitis specimens and normal pancreatic cells, demonstrated overexpression of COX-2 in adenocarcinomas in contrast to non-malignant tissues; additionally, in adenocarcinomas overexpressing COX-2 versus non-expressing ones there was a positive association with severity and poor outcome<sup>[37,38]</sup>. A recent study identified that the COX-2 expression is significantly correlated with the tumor size in pancreatic cancer, which offers a new insight into the regulation of growth and development of metastases in pancreatic

cancer<sup>[39]</sup>. A correlation between COX-2 and p53 expression levels in carcinomas was revealed, and an accumulation of p53 was associated with COX-2 overexpression in premalignant and malignant ductal lesions, which offers opportunities for targeted therapy and chemoprevention of pancreatic cancer using COX-2 inhibitors, especially in lesions with functional p53<sup>[40]</sup>.

### NF-κB

Nuclear factor-kappa B (NF-κB) is a family of cytoplasmic molecules, in normal conditions, which is in inactive status by binding to proteins IκBa and p100. As in many other cancers, NF-κB is activated in pancreatic cancer cells, playing central role in its pathogenesis, progression, invasion and metastasis<sup>[41,42]</sup>. It has been reported that NF-κB overexpression is common in pancreatic cancer. During the last few years a new molecular pathway, the hedgehog (hh) signalling, has been found to play a role in pancreatic carcinogenesis. Inhibition of Shh by cyclopamine (a naturally occurring steroidal alkaloid) resulted in apoptosis induction and blockage of pancreatic cancer cell lines proliferation, both *in vivo* and *in vitro*<sup>[43]</sup>. It is postulated that Shh overexpression is regulated by activation of NF-κB and consequently inhibition of NF-κB results in down-regulation of Shh<sup>[44]</sup>. Another recent study demonstrated that pancreatic cancer cells resistant to gemcitabine exhibit a high basal NF-κB activity<sup>[45]</sup>.

### MMPs

Matrix metalloproteinases (MMPs) are a collection of proteases normally used for tissue remodeling that have been found to be upregulated in the setting of many tumors, including pancreatic cancer. MMP-2 and MMP-9 are likely the most implicated in cancer invasion and metastases among many MMPs<sup>[46]</sup>. As showed in a study by Schneiderhan W et al, pancreatic stellate cells (PSC) represent a significant source of MMP-2 in pancreatic cancer, which is known to promote tumor progression. Moreover, cancer cells release basigin (BSG) to stimulate MMP-2 in PSC, indicating that cross-talk of cancer cells and mesenchymal cells might further increase MMP-2 production in the desmoplasia<sup>[47]</sup>. Moreover, MMP-2 expression correlates with advanced stages of disease and poor prognosis<sup>[48]</sup>. TNF-related apoptosis-inducing ligand (TRAIL) can stimulate the expression of MMP-9 in pancreatic cancer cell lines resulting in the enhancement of invasion involving the signaling pathways of caspases, MEK1/2, PKC and NF-κB<sup>[49]</sup>. Along with

overexpression of specific MMPs in advanced cancer, there is production of the tissue inhibitor of the metalloproteinases (TIMP) which aims to counteract the untoward effects of MMPs in pathological situations<sup>[48]</sup>. The described pathways outline the perplexity of the molecular pathways and their cross talk especially in highly uncontrolled conditions as in malignancies. It seems MMPs play a role as an “angiogenic switch”, to facilitate the expression of proangiogenic factors such as VEGF and bFGF in order to overcome the negative signals of angiogenic inhibitors such as trombospondins, angiostatins, and INFs<sup>[50]</sup>. Due to these properties, the inhibition of MMPs represents the scientific rationale for the development of chemotherapeutic agents against pancreatic cancer<sup>[51]</sup>.

In conclusion, with the comprehensive understanding of its biology, a number of molecularly targeted agents have evolved, which specifically interfere with major pathways involved in pancreatic cancer pathogenesis. Targeted therapy is a promising strategy for pancreatic cancer, where current strategies have failed to significantly improve survival. Furthermore, a biologic and molecular staging of pancreatic cancer may lead us to earlier diagnoses, efficient counseling, and better therapeutic approaches. The ultimate goal would be a tailored-made treatment for maximal therapeutic results.

## REFERENCES

- [1] Jemal A, Siegel R, Ward E, et al. Cancer statistics, 2008[J]. *CA Cancer J Clin* 2008; 58: 71–96.
- [2] Li D, Xie K, Wolff R, et al. Pancreatic cancer[J]. *Lancet* 2004; 363: 1049-57.
- [3] Alexakis N, Halloran C, Raraty M, et al. Current standards of surgery for pancreatic cancer[J]. *Br J Surg* 2004; 91: 1410-27.
- [4] Wray CJ, Ahmad SA, Matthews JB, et al. Surgery for pancreatic cancer: recent controversies and current practice[J]. *Gastroenterology* 2005; 128: 1626-41.
- [5] Oettle H, Post S, Neuhaus P, et al. Adjuvant chemotherapy with gemcitabine vs observation in patients undergoing curative-intent resection of pancreatic cancer: a randomized controlled trial[J]. *JAMA* 2007; 297: 267-77.
- [6] Neoptolemos JP, Stocken DD, Friess H, et al. A randomized trial of chemoradiotherapy and chemotherapy after resection of pancreatic cancer[J]. *N Engl J Med* 2004; 350: 1200-10.
- [7] Pasca di Magliano M, Sekine S, Ermilov A, et al. Hedgehog/Ras interactions regulate early stages of pancreatic cancer[J]. *Genes Dev* 2006; 20: 3161-73.
- [8] van Heek T, Rader AE, Offerhaus GJ, et al. K-ras, p53, and DPC4 (MAD4) alterations in fine-needle aspirates of the pancreas: a molecular panel correlates with and supplements cytologic diagnosis[J]. *Am J Clin Pathol* 2002; 117: 755-65.
- [9] Rozenblum E, Schutte M, Goggins M, et al. Tumor-suppressive pathways in pancreatic carcinoma[J]. *Cancer Res* 1997; 57: 1731-4.
- [10] Ren XY, Liang ZY, Shi XH, et al. A homologous promoterless K-ras CDNA targeting endogenous K-ras expression inhibits human pancreatic cancer cell growth *in vitro* and *in vivo*[J]. *Cancer Biol Ther* 2008; 7: 1641-7.
- [11] Brunner TB, Cengel KA, Hahn SM, et al. Pancreatic cancer cell radiation survival and prenyltransferase inhibition: the role of K-Ras[J]. *Cancer Res* 2005; 65: 8433-41.
- [12] Kawesha A, Ghaneh P, Andren Sandberg A, et al. K-ras oncogene subtype mutations are associated with survival but not expression of p53, p16(INK4A), p21(WAF-1), cyclin D1, erbB-2 and erbB-3 in resected pancreatic ductal adeno- carcinoma[J]. *Int J Cancer* 2000; 89: 469-74.
- [13] Dong M, Ma G, Tu W, et al. Clinicopathological significance of p53 and mdm2 protein expression in human pancreatic cancer[J]. *World J Gastroenterol* 2005; 11: 2162-5.
- [14] Casneuf VF, Fonteyne P, Van Damme N, et al. Expression of SGLT1, Bcl-2 and p53 in primary pancreatic cancer related to survival[J]. *Cancer Invest* 2008; 26: 852-9.
- [15] Sun Y. p53 and its downstream proteins as molecular targets of cancer[J]. *Mol Carcinog* 2006; 45: 409-15.
- [16] Sui X, Shin S, Zhang R, et al. Hdm2 is regulated by K-Ras and mediates p53-independent functions in pancreatic cancer cells[J]. *Oncogene* 2009; 28: 709-20.
- [17] Dong M, Dong Q, Zhang H, et al. Expression of Gadd45a and p53 proteins in human pancreatic cancer: potential effects on clinical outcomes[J]. *J Surg Oncol* 2007; 95: 332-6.
- [18] Schutte M, Hruban RH, Geradts J, et al. Abrogation of the Rb/p16 tumor-suppressive pathway in virtually all pancreatic carcinomas[J]. *Cancer Res* 1997; 57: 3126-30.
- [19] Hu YX, Watanabe H, Ohtsubo K, et al. Frequent loss of p16 expression and its correlation with clinicopathological parameters in pancreatic carcinoma[J]. *Clin Cancer Res* 1997; 3: 1473-7.
- [20] Chen J, Li D, Killary AM, et al. Polymorphisms of p16, p27, p73, and MDM2 modulate response and survival of pancreatic cancer patients treated with preoperative chemoradiation[J]. *Ann Surg Oncol* 2009; 16: 431-9.
- [21] Schulz P, Scholz A, Rexin A, et al. Inducible re-expression of p16 in an orthotopic mouse model of

- pancreatic cancer inhibits lymphangiogenesis and lymphatic metastasis[J]. *Br J Cancer* 2008; 99: 110-7.
- [22] Blobe GC, Schieman WP, Lodish HF, et al. Role of transforming growth factor beta in human disease[J]. *N Engl J Med* 2000; 342: 1350-8.
- [23] Wilentz RE, Iacobuzio Donahue CA, Argani P, et al. Loss of expression of Dpc4 in pancreatic intraepithelial neoplasia: evidence that DPC4 inactivation occurs late in neoplastic progression[J]. *Cancer Res* 2000; 60: 2002-6.
- [24] Tascilar M, Skinner HG, Rosty C, et al. The SMAD4 protein and prognosis of pancreatic ductal adenocarcinoma[J]. *Clin Cancer Res* 2001; 7: 4115-21.
- [25] Cao D, Ashfaq R, Goggins MG, et al. Differential expression of multiple genes in association with MADH4/DPC4/SMAD4 inactivation in pancreatic cancer[J]. *Int J Clin Exp Pathol* 2008; 1: 510-7.
- [26] Iacobuzio-Donahue CA, Fu B, Yachida S, et al. DPC4 gene status of the primary carcinoma correlates with patterns of failure in patients with pancreatic cancer[J]. *J Clin Oncol* 2009; 27: 1806-13.
- [27] Blackford A, Serrano OK, Wolfgang CL, et al. SMAD4 gene mutations are associated with poor prognosis in pancreatic cancer[J]. *Clin Cancer Res* 2009; 15: 4674-9.
- [28] Khorana AA, Hu YC, Ryan CK, et al. Vascular endothelial growth factor and DPC4 predict adjuvant therapy outcomes in resected pancreatic cancer[J]. *J Gastrointest Surg* 2005; 9: 903-11.
- [29] Talar-Wojnarowska R, Malecka-Panas E. Molecular pathogenesis of pancreatic adenocarcinoma: potential clinical implications[J]. *Med Sci Monit* 2006; 12: 186-93.
- [30] Tsiambas E, Karameris A, Dervenis C, et al. HER2/neu expression and gene alterations in pancreatic ductal adenocarcinoma: a comparative immunohistochemistry and chromogenic *in situ* hybridization study based on tissue microarrays and computerized image analysis[J]. *JOP* 2006; 7: 283-94.
- [31] Xiong HQ, Abbruzzese JL. Epidermal growth factor receptor-targeted therapy for pancreatic cancer[J]. *Semin Oncol* 2002; 29(5 suppl 14): 31-7.
- [32] Muslimov GF. Role of epidermal growth factor gene in the development of pancreatic cancer and efficiency of inhibitors of this gene in the treatment of pancreatic carcinoma [J]. *Bull Exp Biol Med* 2008; 145: 535-8.
- [33] Tzeng CW, Frolov A, Frolova N, et al. EGFR genomic gain and aberrant pathway signaling in pancreatic cancer patients[J]. *J Surg Res* 2007; 143: 20-6.
- [34] Azzariti A, Porcelli L, Gatti G, et al. Synergic antiproliferative and antiangiogenic effects of EGFR and mTOR inhibitors on pancreatic cancer cells[J]. *Biochem Pharmacol* 2008; 75: 1035-44.
- [35] Burris H 3rd, Rocha-Lima C. New therapeutic directions for advanced pancreatic cancer: targeting the epidermal growth factor and vascular endothelial growth factor pathways[J]. *Oncologist* 2008; 13: 289-98.
- [36] Chu J, Lloyd FL, Trifan OC, et al. Potential involvement of the cyclooxygenase-2 pathway in the regulation of tumor-associated angiogenesis and growth in pancreatic cancer[J]. *Mol Cancer Ther* 2003; 2: 1-7.
- [37] Juuti A, Louhimo J, Nordling S, et al. Cyclooxygenase-2 expression correlates with poor prognosis in pancreatic cancer[J]. *J Clin Pathol* 2006; 59: 382-6.
- [38] Albazaz R, Verbeke CS, Rahman SH, et al. Cyclooxygenase-2 expression associated with severity of PanIN lesions: a possible link between chronic pancreatitis and pancreatic cancer[J]. *Pancreatology* 2005; 5: 361-9.
- [39] Matsumoto G, Muta M, Tsuruta K, et al. Tumor size significantly correlates with postoperative liver metastases and COX-2 expression in patients with resectable pancreatic cancer[J]. *Pancreatology* 2007; 7: 167-73.
- [40] Hermanova M, Trna J, Nenutil R, et al. Expression of COX-2 is associated with accumulation of p53 in pancreatic cancer: analysis of COX-2 and p53 expression in premalignant and malignant ductal pancreatic lesions[J]. *Eur J Gastroenterol Hepatol* 2008; 20: 732-9.
- [41] Wang Z, Banerjee S, Li Y, et al. Down-regulation of notch-1 inhibits invasion by inactivation of nuclear factor-kappaB, vascular endothelial growth factor, and matrix metalloproteinase-9 in pancreatic cancer cells[J]. *Cancer Res* 2006; 66: 2778-84.
- [42] Holcomb B, Yip Schneider M, Schmidt CM, et al. The role of nuclear factor kappaB in pancreatic cancer and the clinical applications of targeted therapy[J]. *Pancreas* 2008; 36: 225-35.
- [43] Thayer SP, di Magliano MP, Heiser PW, et al. Hedgehog is an early and late mediator of pancreatic cancer tumorigenesis[J]. *Nature* 2003; 425: 851-6.
- [44] Nakashima H, Nakamura M, Yamaguchi H, et al. Nuclear factor-kappaB contributes to hedgehog signaling pathway activation through sonic hedgehog induction in pancreatic cancer[J]. *Cancer Res* 2006; 66: 7041-9.
- [45] Scwab GM, Fujioka S, Schmidt C, et al. NF-kappaB in pancreatic cancer[J]. *Int J Gastrointest Cancer* 2003; 33: 15-26.
- [46] Gong YL, Xu GM, Huang WD, et al. Expression of matrix metalloproteinases and the tissue inhibitors of metalloproteinases and their local invasiveness and metastasis in Chinese human pancreatic cancer [J]. *J Surg Oncol* 2000; 73: 95-9.

- [47] Schneiderhan W, Diaz F, Fundel M, et al. Pancreatic stellate cells are an important source of MMP-2 in human pancreatic cancer and accelerate tumor progression in a murine xenograft model and CAM assay[J]. *J Cell Sci* 2007; 120: 512-9.
- [48] Juuti A, Lundin J, Nordling S, et al. Epithelial MMP-2 expression correlates with worse prognosis in pancreatic cancer[J]. *Oncology* 2006; 71: 61-8.
- [49] Zhou DH, Trauzold A, Röder C, et al. The potential molecular mechanism of overexpression of uPA, IL-8, MMP-7 and MMP-9 induced by TRAIL in pancreatic cancer cell[J]. *Hepatobiliary Pancreat Dis Int* 2008; 7: 201-9.
- [50] Rundhaug JE. Matrix metalloproteinases, angiogenesis, and cancer: commentary re: A. C. Lockhart et al, Reduction of wound angiogenesis in patients treated with BMS-275291, a broad spectrum matrix metalloproteinase inhibitor[J]. *Clin Cancer Res* 2003; 9: 551-4.
- [51] McCawley LJ, Matrisian LM. Matrix metalloproteinases: they're not just for matrix anymore[J]. *Curr Opin Cell Biol* 2001; 13: 534-40.